



Microbiome workshop – data generation and processing

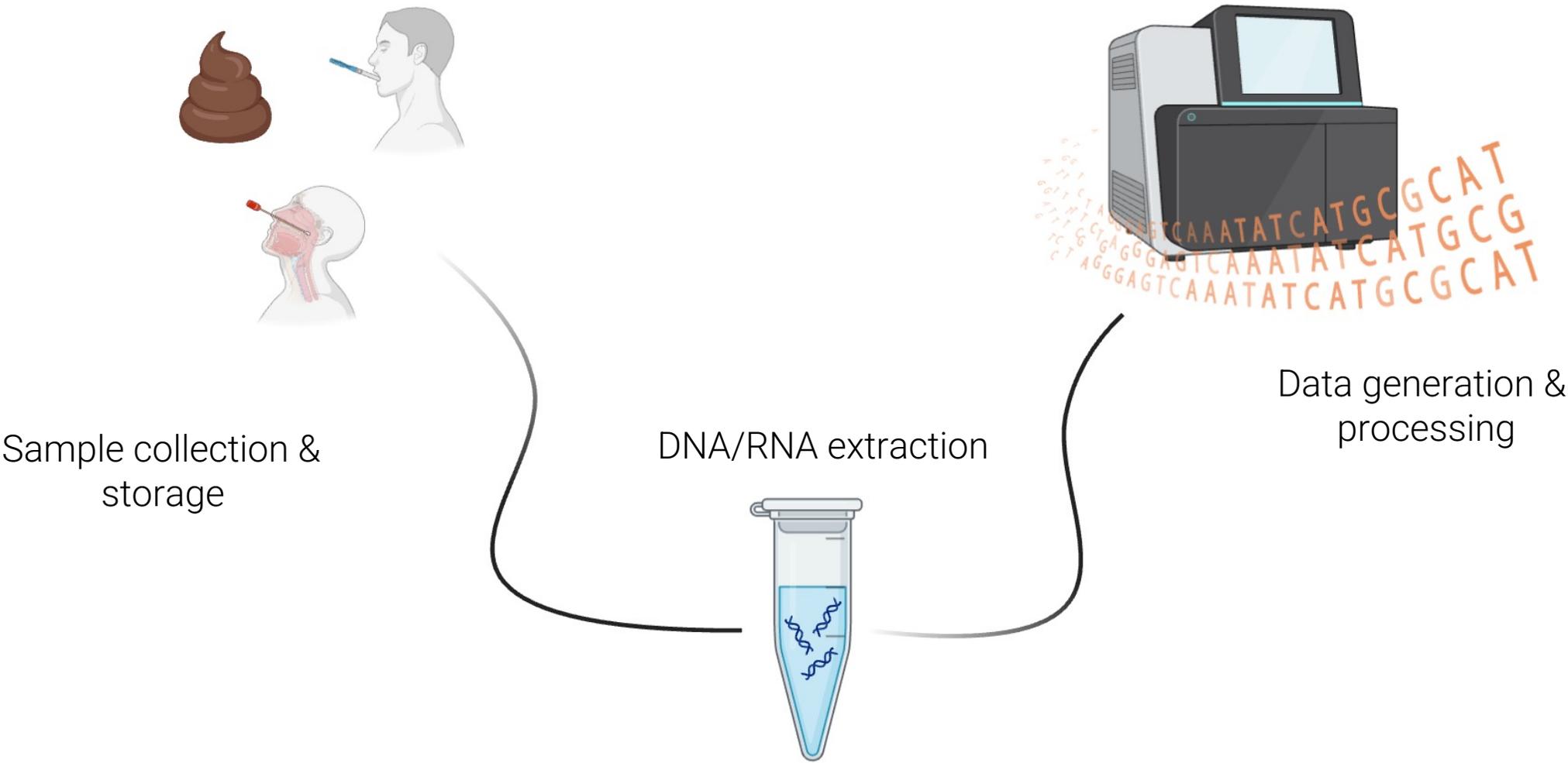
September 2024



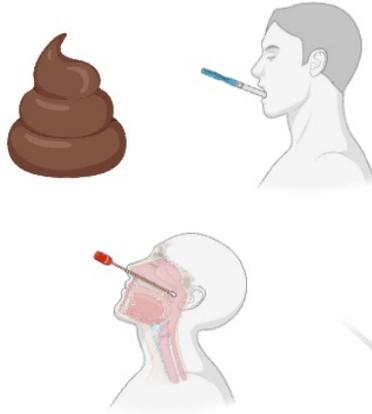
IKMB

Institute of Clinical
Molecular Biology

Microbiome science - from sample to data



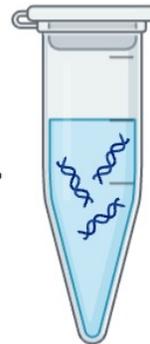
Microbiome science - from sample to data



Sample collection & storage

- Sampling method (swab, tube, ...)
- Sampling buffer (RNAlater, glycerol, ...)
- Storage conditions (snap freezing, -20/-80°C)

DNA/RNA extraction



- Extraction kit used
- DNA or RNA or both?
- Cell disruption (Proteinases, Bead beating)

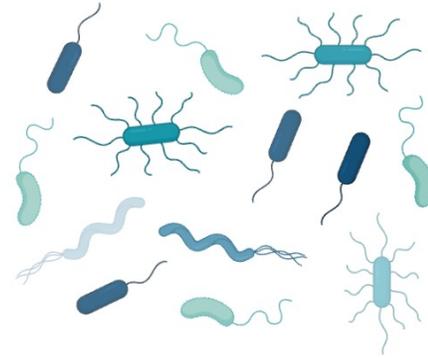


Data generation & processing

- 16S rRNA gene or Shotgun metagenomics
- Short reads or long reads
- Data annotation

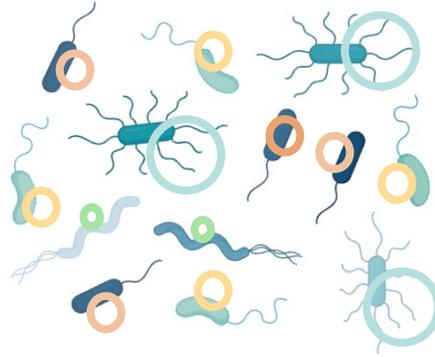
16S rRNA gene
amplicon sequencing

shotgun
metagenomics



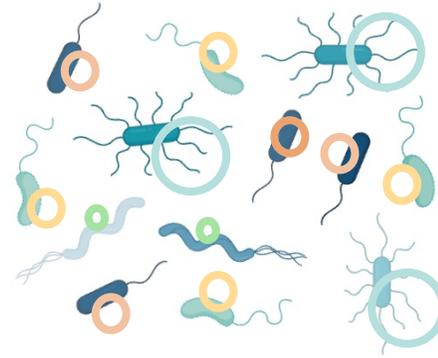
16S rRNA gene
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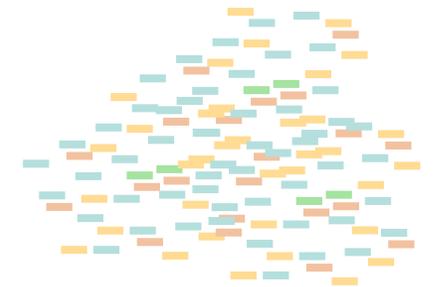
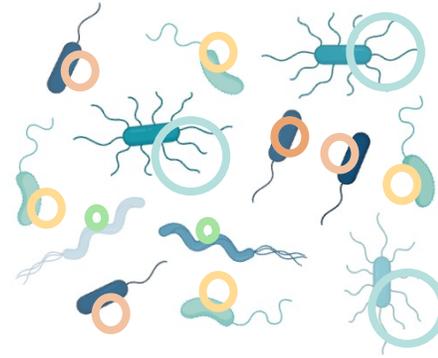
16S rRNA gene amplicon sequencing

shotgun metagenomics



16S rRNA gene amplicon sequencing

shotgun metagenomics



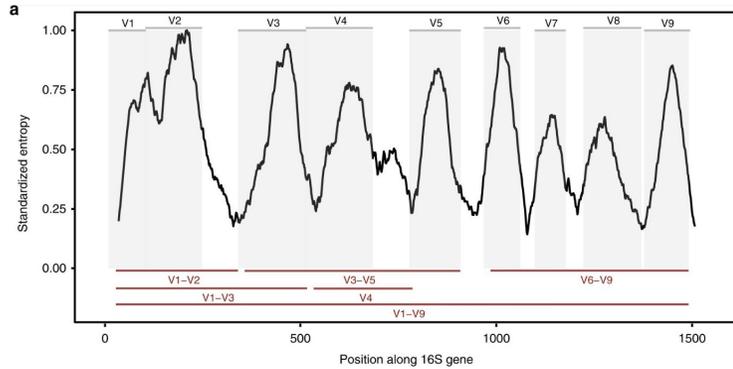
Full genome information + + +

More data + / -

More expensive - -

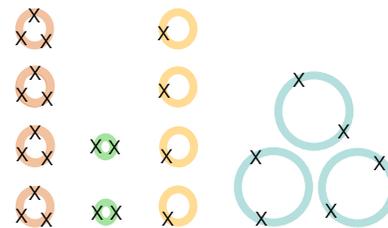
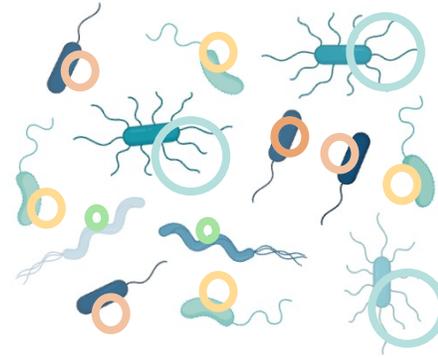
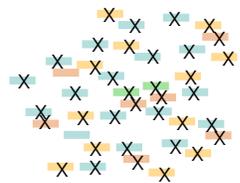
Bias towards high abundant taxa -

16S rRNA gene amplicon sequencing



16S rRNA gene: conserved and variable regions

PCR target + Molecular clock = taxon information



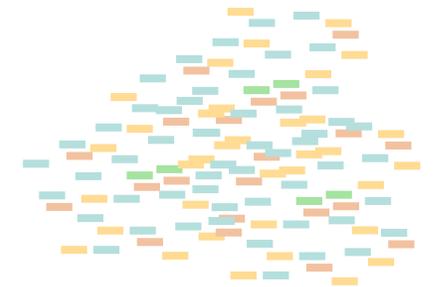
shotgun metagenomics

Full genome information + + +

More data + / -

More expensive - -

Bias towards high abundant taxa -



What we have: The FastQ format

Identifier | @HWI-EAS209_0006_FC706VJ:5:58:5894:21141#ATCACG/1
Sequence | TTAATTGGTAAATAAATCTCCTAATAGCTTAGATNTTACCTTNNNNNNNNNNNTAGTTTCTTGAGA
+ sign & identifier | +HWI-EAS209_0006_FC706VJ:5:58:5894:21141#ATCACG/1
Quality scores | efcfffffcfeefffcfffffddf`feed]`]_Ba_^__ [YBBBBBBBBBBRTT\]] [] dddd`

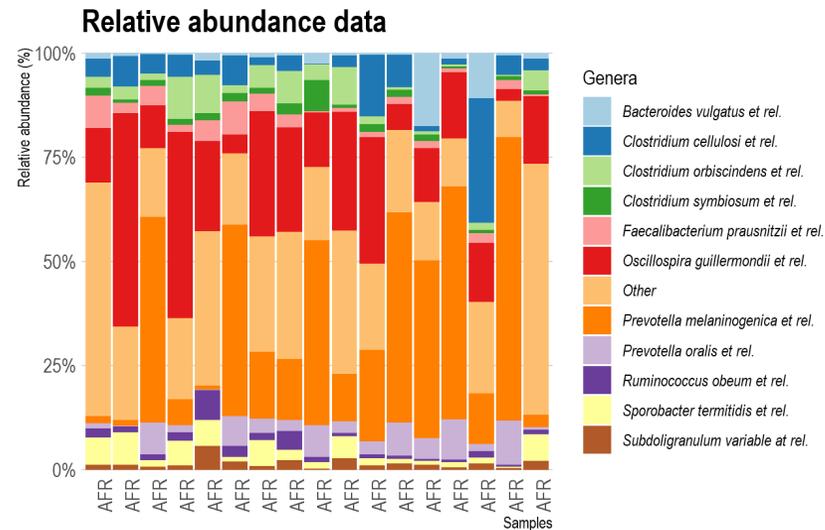
Base T
phred Quality] = 29

What we have: The FastQ format

Identifier | @HWI-EAS209_0006_FC706VJ:5:58:5894:21141#ATCACG/1
Sequence | TTAATTGGTAAATAAATCTCCTAATAGCTTAGATNTTACCTTNNNNNNNNNTAGTTTCTTGAGA
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Quality scores | efcfffffcfeefffcfffffdff`feed]`_]_Ba_^__[YBBBBBBBBBRTT\]][] dddd`

Base T
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What we want:



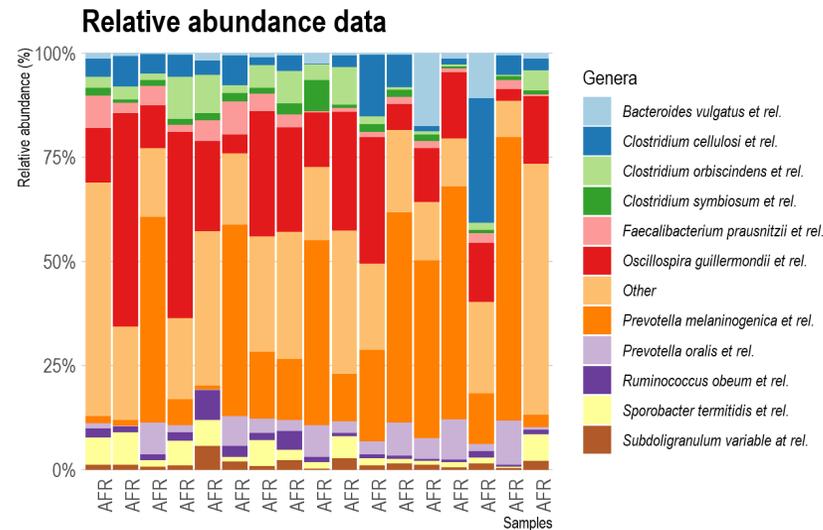
<https://compgenomr.github.io/book/fasta-and-fastq-formats.html>

What we have: The FastQ format

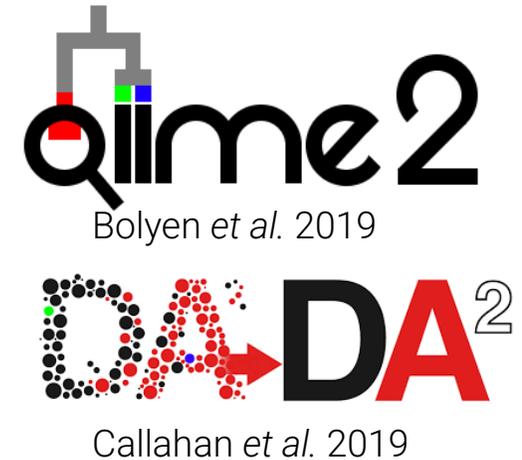
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+ sign & identifier | +HWI-EAS209_0006_FC706VJ:5:58:5894:21141#ATCACG/1  
Quality scores | efcfffffcfeefffcfffffddf`feed]`_]_Ba_^__[YBBBBBBBBBRTT\]] [ ] dddd`
```

Base T
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What we want:



How we get there:



<https://compgenomr.github.io/book/fasta-and-fastq-formats.html>

16S data workflow with



Version: 2024.5 ▾

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Quick search

QIIME 2 user documentation

This site is the official user documentation for QIIME™ 2, including installation instructions, tutorials, and other important information. Visit <http://qiime.org> for information on QIIME™ 1.

Getting started

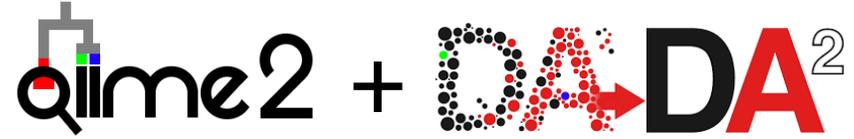
Check out the [getting started](#) guide to begin using QIIME 2.

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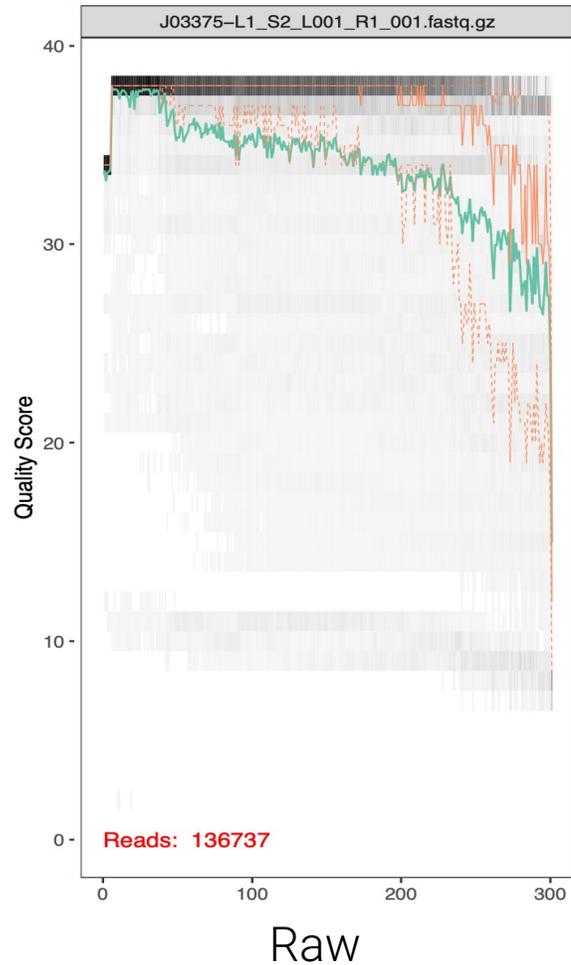
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 - [QIIME 2 for Experienced Microbiome Researchers](#)

- **Tutorials**
 - Overview of QIIME 2 Plugin Workflows
 - QIIME 2 for Experienced Microbiome Researchers
 - “Moving Pictures” tutorial
 - “Moving Pictures” tutorial - Multiple Interface Edition
 - Fecal microbiota transplant (FMT) study: an exercise
 - “Atacama soil microbiome” tutorial
 - Parkinson’s Mouse Tutorial
 - Importing data
 - Exporting data
 - Metadata in QIIME 2
 - Filtering data
 - Training feature classifiers with q2-feature-classifier
 - Evaluating and controlling data quality with q2-quality-control
 - Predicting sample metadata values with q2-sample-classifier
 - Performing longitudinal and paired sample comparisons with q2-longitudinal
 - Identifying and filtering chimeric feature sequences with q2-vsearch
 - Alternative methods of read-joining in QIIME 2
 - Clustering sequences into OTUs using q2-vsearch
 - Utilities in QIIME 2
 - Phylogenetic inference with q2-phylogeny

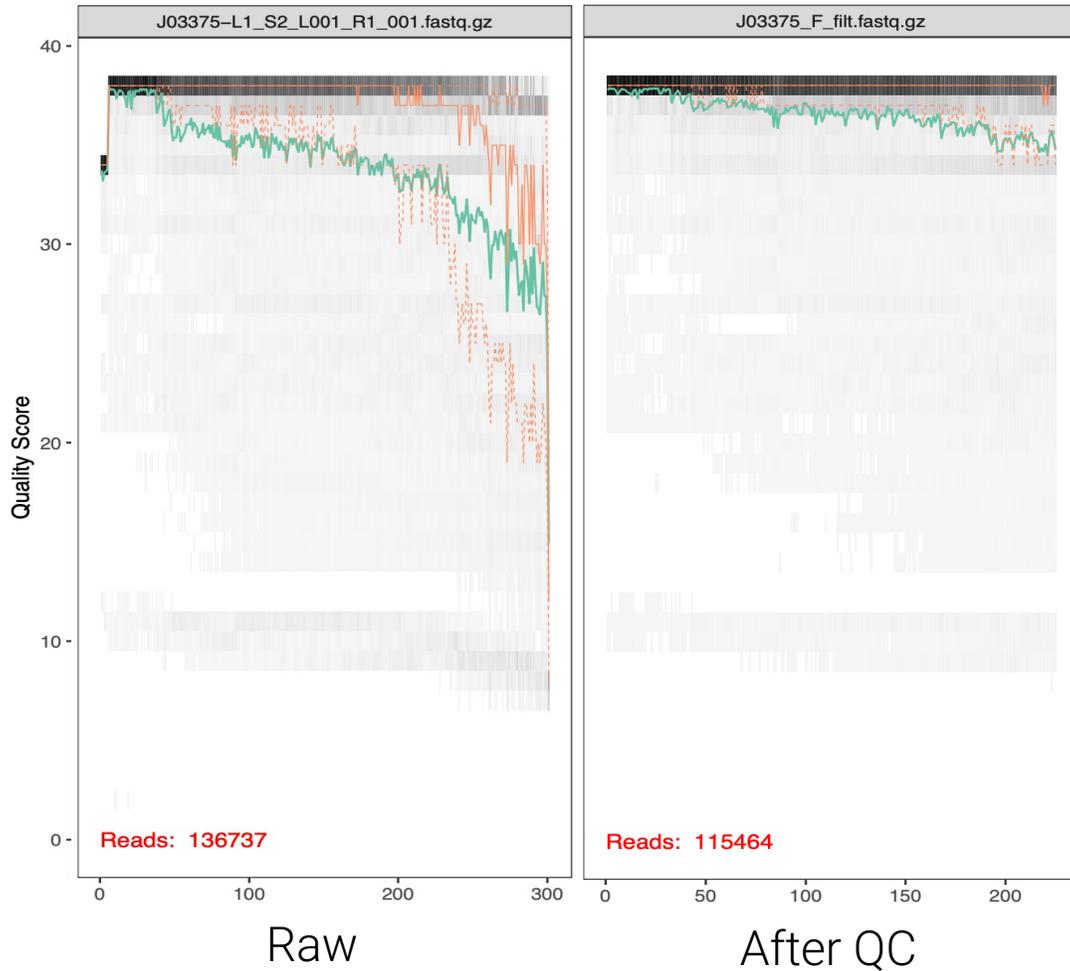
16S data workflow with



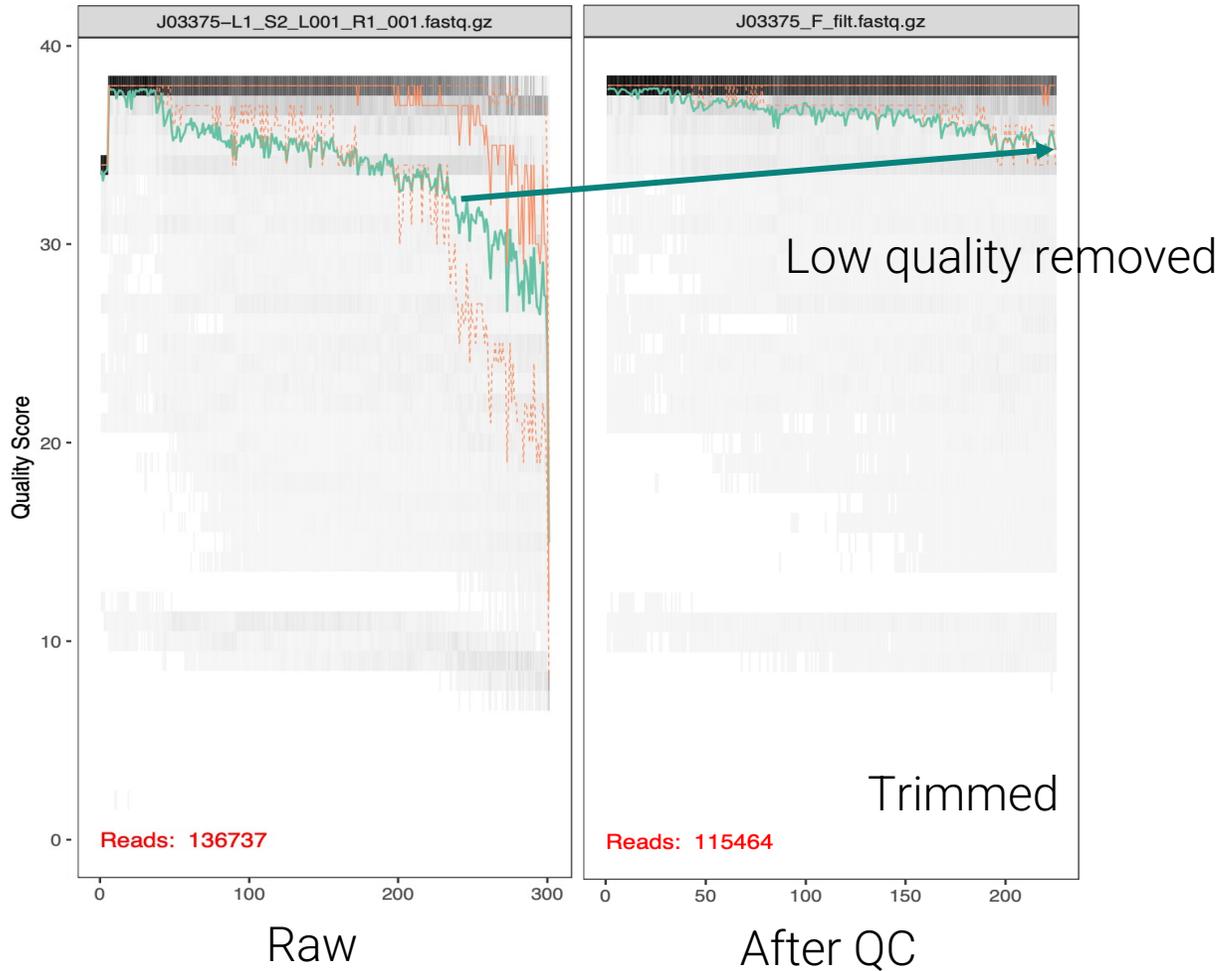
Step 1: QC and trimming



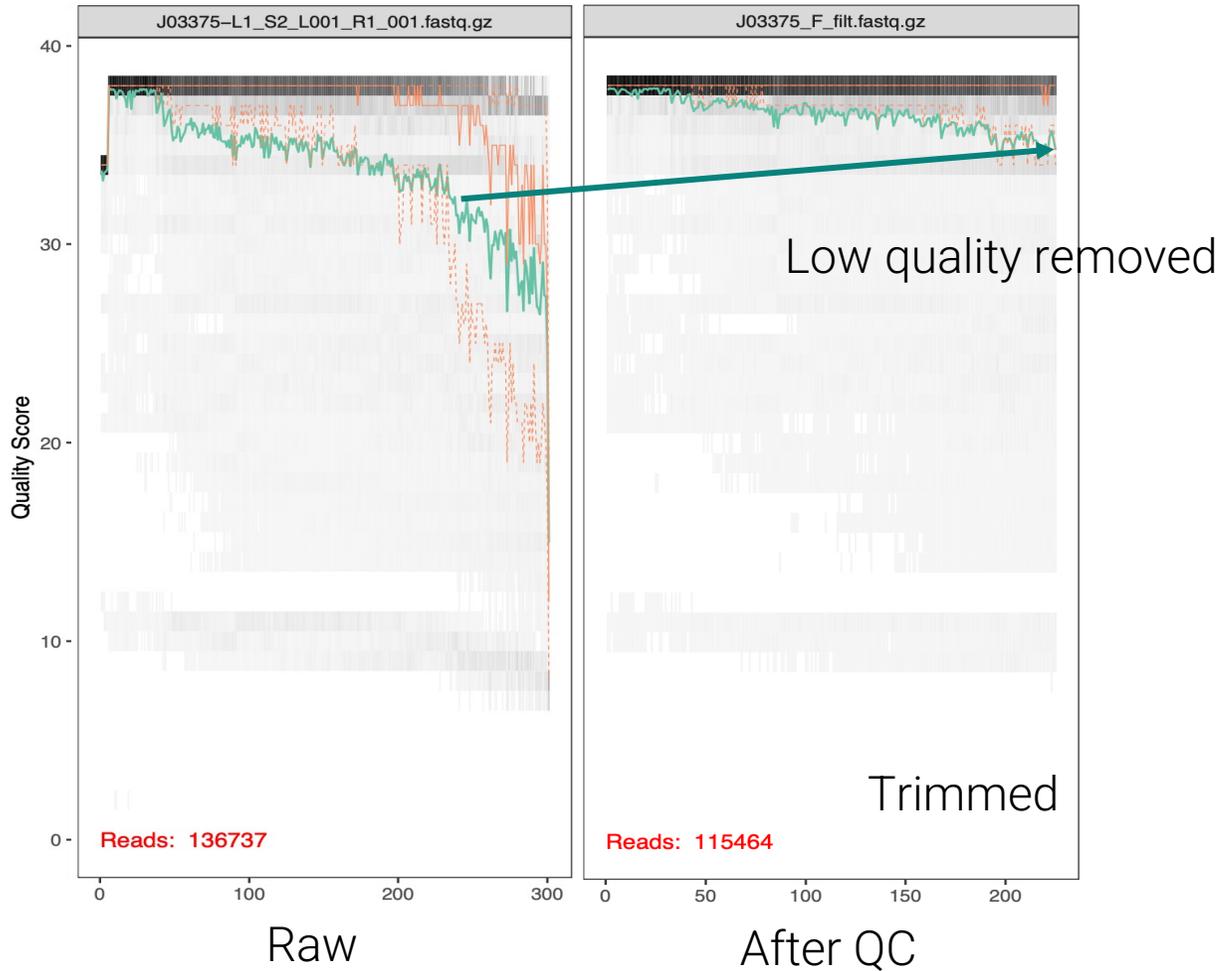
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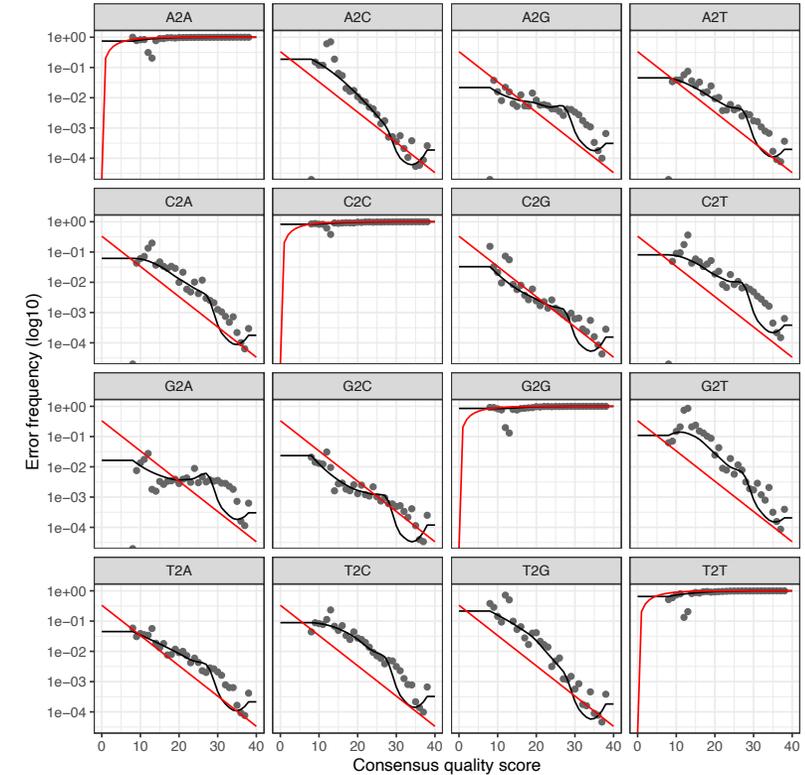
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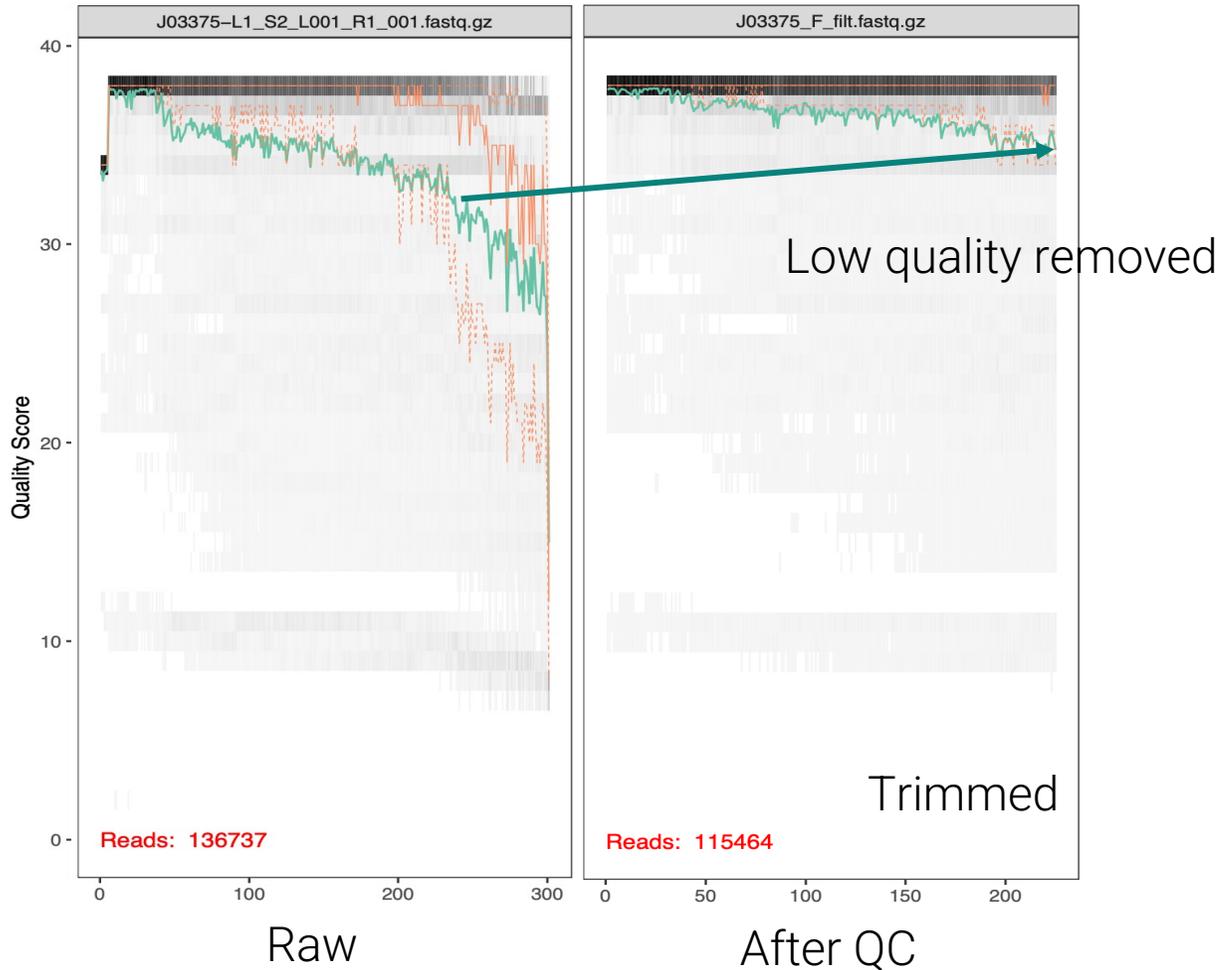
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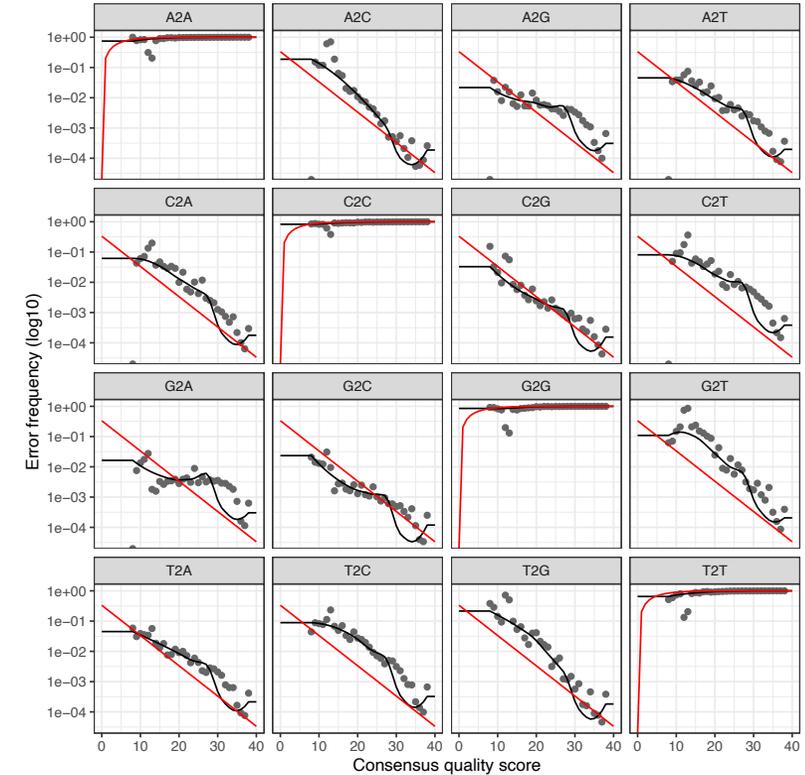
Step 2: Error correction



Step 1: QC and trimming



Step 2: Error correction



Step 3:
Inference of Amplicon Sequence Variants (ASVs)

Assigning taxonomic labels

Abundance Table

	ASV1	ASV2	ASV3	ASV4
S1	5	10	3	7
S2	5	0	3	0
S3	0	0	4	3

ASV sequences:

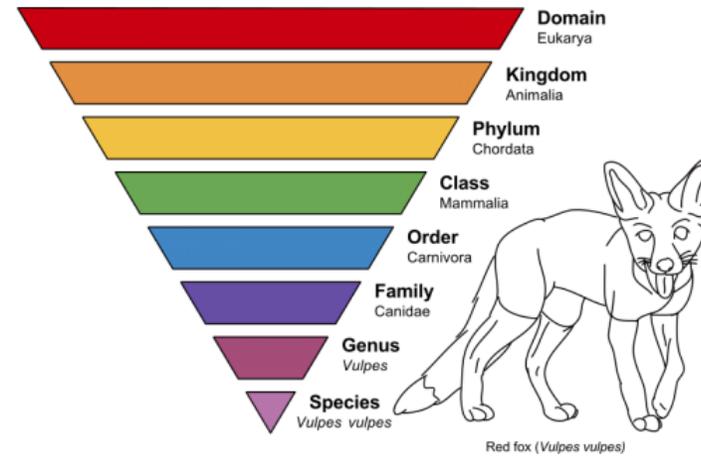
```
>ASV1
ACGCTGGCGGCATGCTTAACACATGCAAGTCGTACGAATGAAT...
>ASV2
ACGCTGGCGGTATGCCTAACACATGCAAGTCGAACGAGGTAGC...
>ASV3
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAACAG...
>ASV4
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAGTCA...
```

Assigning taxonomic labels

Abundance Table

	ASV1	ASV2	ASV3	ASV4
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What is a taxonomy?



ASV sequences:

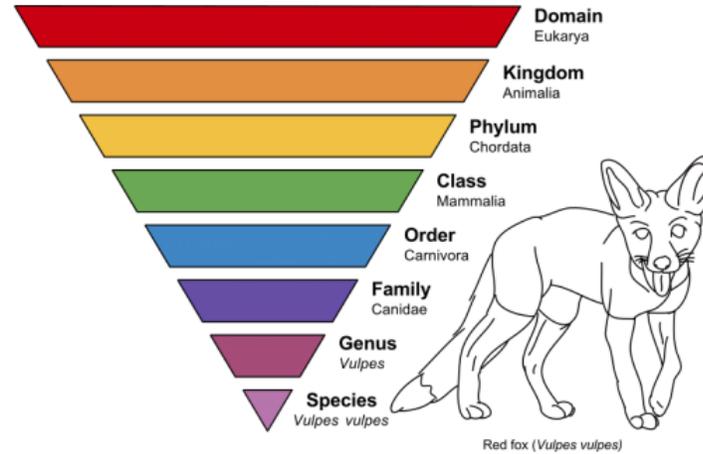
```
>ASV1
ACGCTGGCGGCATGCTTAACACATGCAAGTCGTACGAATGAAT...
>ASV2
ACGCTGGCGGTATGCCTAACACATGCAAGTCGAACGAGGTAGC...
>ASV3
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAACAG...
>ASV4
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```

Assigning taxonomic labels

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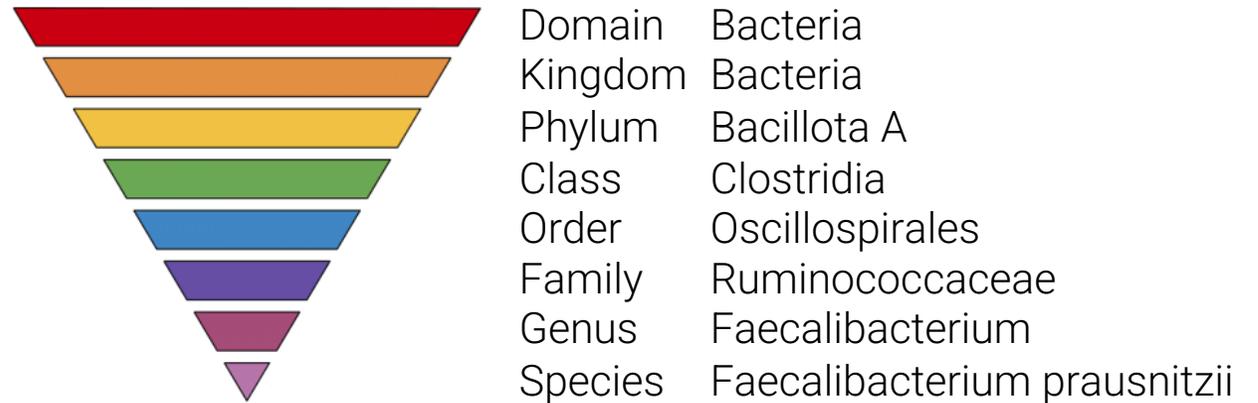
What is a taxonomy?



ASV sequences:

```
>ASV1
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>ASV2
ACGCTGGCGGTATGCCTAACACATGCAAGTCGAACGAGGTAGC...
>ASV3
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAACAG...
>ASV4
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAGTCA...
```

Same can be done for microorganisms:



Assigning taxonomic labels

Abundance Table

	ASV1	ASV2	ASV3	ASV4
S1	5	10	3	7
S2	5	0	3	0
S3	0	0	4	3

ASV sequences:

```
>ASV1
ACGCTGGCGGCATGCTTAACACATGCAAGTCGTACGAATGAAT...
>ASV2
ACGCTGGCGGTATGCCTAACACATGCAAGTCGAACGAGGTAGC...
>ASV3
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAACAG...
>ASV4
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAGTCA...
```

Database search
(Naive Bayesian Classifier)



Assigning taxonomic labels

Abundance Table

	ASV1	ASV2	ASV3	ASV4
S1	5	10	3	7
S2	5	0	3	0
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ASV sequences:

```
>ASV1
ACGCTGGCGGCATGCTTAACACATGCAAGTCGTACGAATGAAT...
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ACGCTGGCGGTATGCCTAACACATGCAAGTCGAACGAGGTAGC...
>ASV3
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAACAG...
>ASV4
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAGTCA...
```



Greengenes2



Ribosomal database project



Database search
(Naive Bayesian Classifier)

Assigning taxonomic labels

Abundance Table

	ASV1	ASV2	ASV3	ASV4
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ASV sequences:

```
>ASV1
ACGCTGGCGGCATGCTTAACACATGCAAGTCGTACGAATGAAT...
>ASV2
ACGCTGGCGGTATGCCTAACACATGCAAGTCGAACGAGGTAGC...
>ASV3
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAACAG...
>ASV4
ACGCTAGCGGCAGGCTTAACACATGCAAGTCGAGGGGTAGTCA...
```



Greengenes2



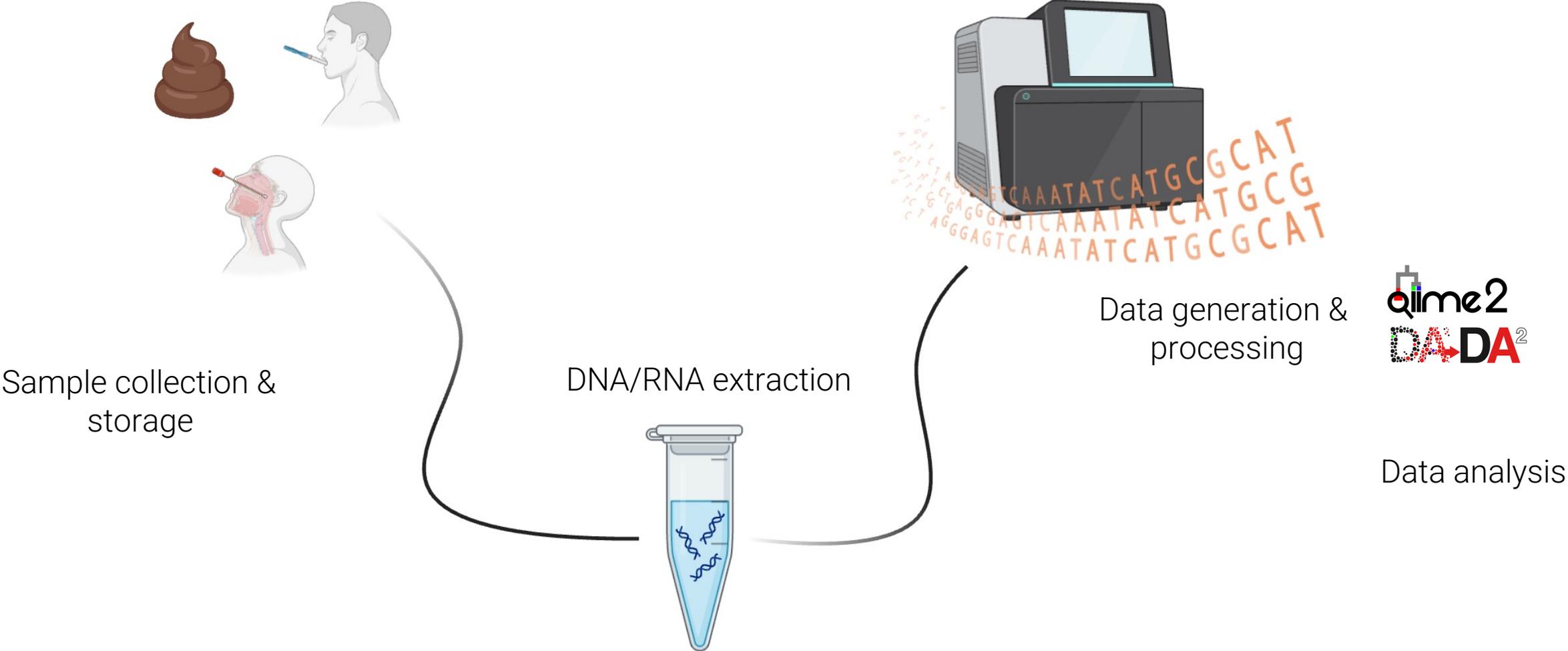
Ribosomal database project



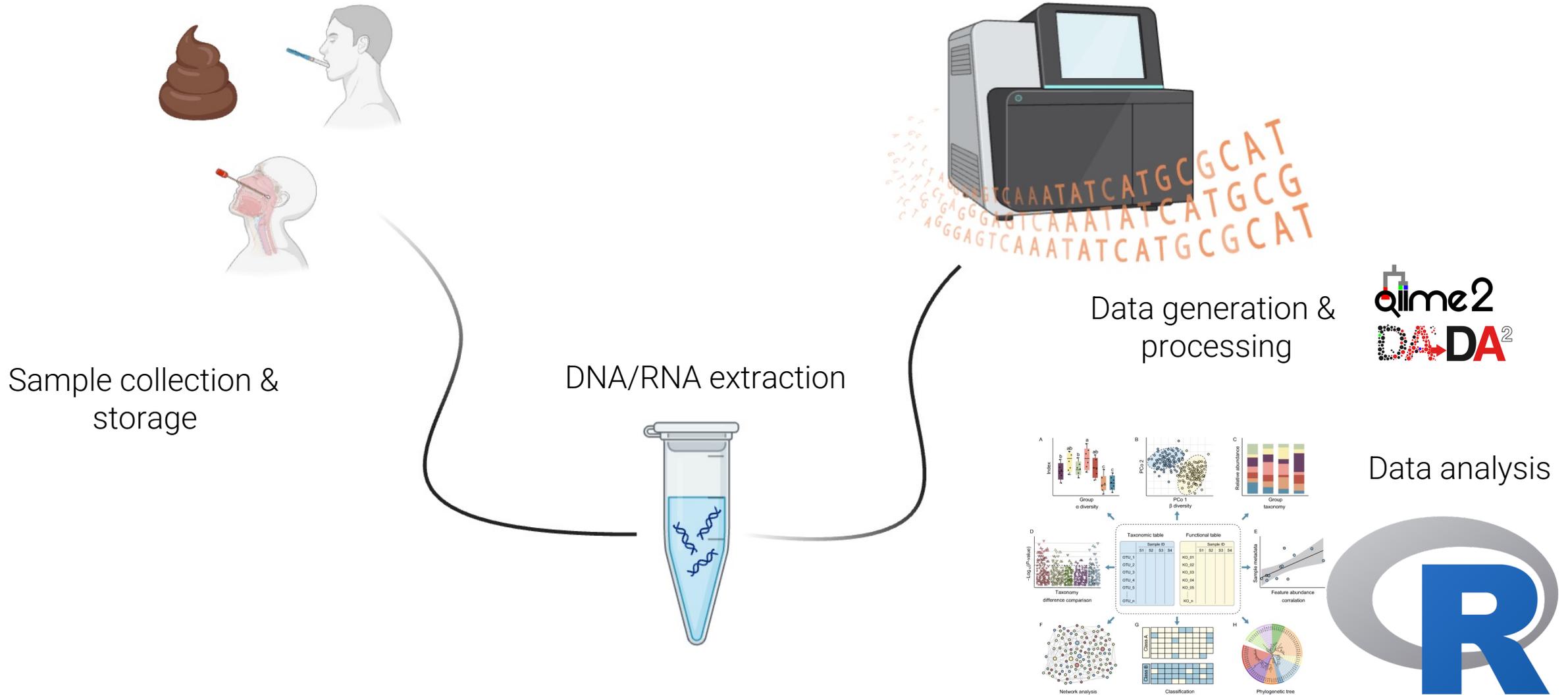
Database search
(Naïve Bayesian Classifier)

	Domain	Phylum	Class	Order	Family	Genus
ASV1	Bacteria	Bacillota_C	Negativicutes	Veillonellales	Dialisteraceae	Dialister
ASV2	Bacteria	Bacteroidota	Bacteroidia	Bacteroidales	Bacteroidaceae	Bacteroides
ASV3	Bacteria	Pseudomonadota	Gammaproteo bacteria	Enterobacterales	Enterbacteriaceae	
ASV4	Bacteria	Bacteroidota	Bacteroidia	Bacteroidales	Bacteroidaceae	Prevotella

Microbiome science - from sample to data



Microbiome science - from sample to data to analysis



RStudio

Project: (None)

AlphaDiversity.R x Untitled3* x Intro_R.R x 1_IgA_CoatingIndexCalculation_OT. >>

Source on Save Run Source

```

1
2 ## My microbiome analyses
3
4 library(phyloseq)
5
6 setwd("/Users/apschaan/Documents/Kiel/UKSH-CAU/Classes/KMC_workshop")
7
8 taxTable <- read.csv("taxonomy_gmbc1_form.csv", header=TRUE, row.names=1)
9
10 head(taxTable)
11
12
13
14

```

Environment History Connections

Global Environment Taxa

Data

- TaxaForHeatm... 40 obs. of 12 variables
- TaxaForHeatm... 20 obs. of 2 variables

Values

taxa	"a361d6b053ce9d38182128a316635e3e"
taxa_names	chr [1:197] "2e63e8e3f9f0f102f570e7..."
taxaKeep	chr [1:2174] "9492bc05eee3b253c1557..."

Values

taxa	"a361d6b053ce9d38182128a316635e3e"
taxa_names	chr [1:197] "2e63e8e3f9f0f102f570e7..."
taxaKeep	chr [1:2174] "9492bc05eee3b253c1557..."

Files Plots Packages Help Viewer Presentation

Zoom Export

Wilcoxon, p = 4.3e-05

log(|C|)

Industrialized Non-industrialized

Console Terminal Background Jobs

```

R 4.3.1 ~ /Documents/Kiel/UKSH-CAU/Classes/KMC_workshop/
> library(phyloseq)
> setwd("/Users/apschaan/Documents/Kiel/UKSH-CAU/Classes/KMC_workshop")
> taxTable <- read.csv("taxonomy_gmbc1_form.csv", header=TRUE, row.names=1)
> head(taxTable)

```

	Kingdom	Phylum
675c847bccbc53942ebb7b8cbb4efc4d	d__Bacteria	p__Bacteroidota
da6a7cba87e0895b9cbe6037b9bd8b3b	d__Bacteria	p__Bacteroidota